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(84) **HIV-2 virus variants.**

(57) **HIV-2 virus variants, namely virus HIV D205, which can be cloned from the corresponding virus isolate HIV D205 (ECACC V 87122304) and its RNA or RNA-fragments and DNA and DNA-fragments derived therefrom and/or proteins and the use thereof for diagnostics and therapy.**

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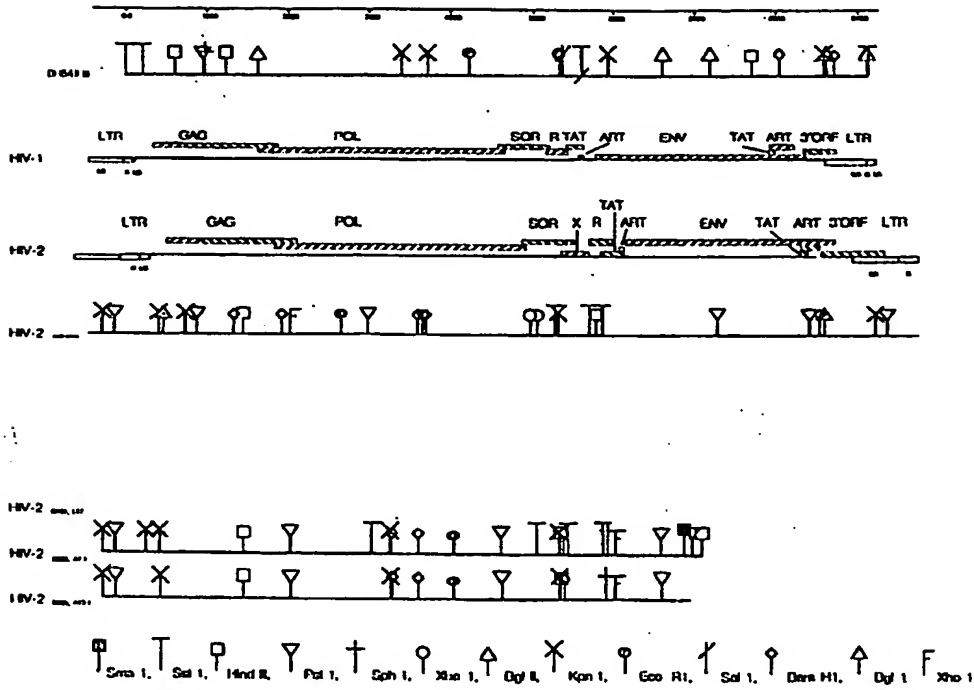


Fig. 1

The present invention relates to HIV D205 a HIV-2 virus variant that may be cloned from the corresponding virus isolate HIV D205 (ECACC V 87122304).

"Molecular cloning of two West African human immunodeficiency virus type 2 isolates which replicate well on macrophages: a Gambian isolate from a case of neurologic acquired immunodeficiency syndrome, and a highly divergent Ghanesian isolate" (Kühnel, H., v. Briesen, H., Dietrich, U., Adamski, M., Mix, D., Biesert, L. Kreutz, R., Immelmann, A., Henco, K., Meichsner, Ch., Andreessen, R., Gelderblom, H. & Rübsamen-Waigmann, H., 1989, Proc. Natl. Acad. Sci. 86, 4, 2383-2387.

In diagnostics, two criteria are demanded to be met, namely specificity and sensitivity for the antigen to be detected. In the diagnostics of AIDS the demand for specificity can certainly be complied with by using the isolates HTLV-III_B and LAV-2 (Guyader, M. et al., "Nature" 326, 1987, 662-669) in order to delimit HIV infections from other infections and, thus, to make a rough assignment into the classes of "HIV-2-related infections" or "HIV-1-related infections". However, a problem is constituted by the sensitivity of the diagnosis. In the range of the so-called seroconversion, i.e. the initial occurrence of the antibody in the infected person, a reduction in sensitivity implies an increase in the number of "falsely negative" test results. Accordingly, it is one main goal to shorten the period between an infection and the detectability of this infection as much as possible by improving the test sensitivity.

A decreased cross reactivity, in the practice of the widely employed ELISA diagnostics, is manifested, for example, in a reduced sensitivity. Thus, the use of the described HIV-1 isolate means about an average reduction of the test sensitivity against HIV-2 sera by the factor of 100 to 1000, whereas the isolate HTLV-III_B enables almost no detection to be accomplished anymore.

A disastrous principle of the diseases caused by HIV resides in the fact that there is not only one type of each of HIV-1 and HIV-2 virus phenotypes and genotypes. What is to be premised is rather a large group of related viruses, possible even populations which by no way are strictly separated from each other but continuously penetrate one another and undergo some evolutionary development to a more and more increasing divergence, while at the same time they begin by recombination events to exchange between each other parts of the genom. Thus, the existing HIV species form a broad continuous population level in which there are no narrowly delimited subpopulations of one virus variant. There is rather to presumed that a continuum exists which is subject to permanent fluctuations with time.

The classified virus variants HIV-1 and HIV-2 are representatives of the diffusely delimited subpopulations having a relative low degree of relationship, which is manifested by only a partial cross reactivity. On the other hand, there are variants of the HIV-1 group (Rübsamen-Waigmann, H. et al., "AIDS-Forschung" 10, 1987, 572-575; Rübsamen-Waigmann, H. et al., J. Med. Virol. 19, 1986, 335-344; v. Briesen, H. et al., J. Med. Virol. 23, 1987 51-66), which do significantly stronger cross-react with HIV-2 than the first characterized HIV-1 isolate itself (Hahn, B. et al., "Nature" 312, 1984, 166-169). A commercial product consisting of such an isolate diagnoses distinctly more sera as being HIV-2 positive than does the described standard isolate HTLV-III_B.

An ideal diagnostic or therapeutic product should contain at least one representative from the populations as significantly biologically distinguished from one another.

HIV-1 viruses in a multitude of highly polymorphic genetic mutants may cause different diseases such as ARC, LAS, AIDS and encephalopathies (ARC: AIDS-related complex, LAS: lymphadenopathy syndrome, AIDS acquired Immune deficiency syndrome). Cloned virus variants are distinguished in sequence and restriction pattern, even if they have been isolated at the same time, at the same place and even from the same patient (Rübsamen, H. et al., 1986). It could be shown that virus variants of the HIV-1 type are distinguished in some virus antigens up to about 15%. HIV-2's are even different in more than 40% of the aminoacids in some antigens, substitutions, insertions and deletions having been considered (Guyader, M. et al., 1987; Rabson, A.B. & Martin, M.A. "Cell" 40, 1985, 477-480).

The present invention provides a variant of the HIV-2 virus. The variant was isolated from a clinically asymptomatic patient. The virus isolate proved to be diagnostic agents, relative to DNA/RNA as well as relative to the virus antigens, for serologically and directly identifying infections by the type HIV-2 in the pre-AIDS and AIDS stages.

The virus isolate according to the invention comprises viruses and proviruses, the characteristics of which are identical to those of the disclosed restriction map and the sequence of the cloned partial regions (Figures 1-4). Moreover, the virus isolate comprises variants which are distinguished from the viruses and proviruses described above in that they are different in their nucleotide sequences from the above-described viruses only by up to 5%, and preferably by 2%, particularly preferred by 1%.

The virus variant according to the invention may cause lymphadenopathies (further designated as LAS/AIDS). Claimed according to the invention are also expression products of said virus variant, and more particularly antigens, preferably in accumulated or pure form, and processes for producing said expression

products in full or in parts or in combinations of the parts. The expression products are intended to include all polypeptides in glycosylated and/or meristylated forms which have been coded on the positive or negative strand of the cloned RNA or DNA.

A further preferred embodiment consists of cloned DNA sequences capable of hybridizing with genomic RNA and DNA of the virus variant. Claimed according to the invention are stable gene probes containing such DNA sequences which are suitable for the detection of hybridization of those and other HIV variants or related viruses or DNA proviruses in samples to be investigated, more particularly biological or semi-synthetic samples.

A further preferred embodiment of the invention is comprised by virus variant the RNA/DNA of which or respective fragments will hybridize to the virus variants according to the invention under stringent conditions, more particularly c-DNA, genomic DNA, recombinant DNA, synthetic DNA or fragments thereof. These are understood to include variants or fragments which exhibit deletions and insertions in comparison to the virus variant according to the invention.

Stringent conditions of hybridization and washing are meant to be understood as those conditions which ensue by way of experiment or calculation if the melting point of the 100% homologous nucleic acid complexes in conditions of hybridization and washing will be fallen below by not more than 5 °C under the buffer conditions employed.

Also claimed according to the invention are cloned synthetic gene probes which may be derived from the above-described virus variants and can be augmented in vector systems in eukaryotes or prokaryotes. The described cloned DNA fragments are suitable for hybridization with complementary nucleic acids (DNA/RNA) for the purpose of diagnostic detection of the virus variants. The diagnostic tests according to the invention are carried out by using DNA or RNA probes. The probes are radioactive or have been labelled with fluorescent bio- or chemiluminescent groups or enzymes or are specifically detectable with enzymes via coupled reaction systems. The hybridizations may be effected in a homogeneous phase of a solution or in a heterogeneous phase with solid-immobilized nucleic acids, while the solid may be a membrane, particle, cell or tissue, so that the hybridization may also be effected *in situ*.

From the virus isolate claimed according to the invention, the corresponding DNA sequences (Figure 1) may be cloned in *E. coli* bacteria by establishing a genomic lambda-gene bank, starting from the DNA of the lymphocytes infected with the virus isolate. The desired clones are obtained by carrying out a plaque-screening with STLIV-III sequences of the gag-pol range. In a more specific way, there may be used as a probe a DNA derived from the published sequence HIV-2 ROD (Guyader, M. et al., "Nature" 326, 1987, 662-669), or a DNA probe derived from the partial sequences of the isolate HIV-2 D205 according to the invention.

The diagnostic method based on the use of the viruses claimed according to the invention comprises the following steps: Extraction of RNA or DNA from biological samples, possibly enzymatic processing by restriction enzymes, separation by gel electrophoresis and/or direct blot methods for nucleic acid-binding carriers, and subsequent hybridization with parts of the cloned fragments of the claimed viruses. Hybridizations may also be directly carried out in chemically treated cells or tissues. Therein the origin of the tissues or liquids is insignificant.

Specifically, a process for the *in vitro* detection of antibodies against expression products of the viruses of the present invention is characterized in that the expression products or parts thereof of the viruses are detected by means of immunological methods. The process is characterized in that the expression products are proteins, peptides or parts thereof which have been coded within the meaning of an open reading frame on the DNA of the proviral partial sequences as characterized in claim 1 and are prepared by synthetic or biosynthetic processes.

The process is further characterized in that previously a definite amount or a combination of expression products or parts thereof are fixed on microtiter plates, whereupon subsequently biological samples, diluted or undiluted, are contacted with the coated microtiter plates and after incubation and sequential washing steps can be identified by means of a detecting reagent or of labelled anti-HIV antibodies.

Alternatively, filter strips and plastic strips or rods are used instead of microtiter plates, wherein the expression products of the viruses have been fixed at respective specific positions by isolated application of the different antigens.

The expression products or parts thereof can also be separated by gel electrophoresis and then transferred by blotting whereupon incubation with anti-HIV antibodies and the detection thereof are effected. Detection is effected on solid phase carriers to which the antigen determinants have been bonded, with the solid phase carrier consisting of particles.

Expression products can be virus antigens derived from *in vitro*-infected cells, said antigens being contacted with biological test materials as antigens bonded to fixed cells, and that the subsequent antibody

bonding can be determined with immunological detection reagents by means of an apparatus, for example with a cytofluorimeter, or visually.

The antigens can be determined by competitive ELISA. HIV-related nucleic acids (DNA and RNA) can be detected in biological samples, cells and in isolated form by using the nucleic acids according to the present invention.

Expression products can be supplemented by materials which are related to other HIV variants, which, however, are distinguished in their biological properties from the materials of the isolates of the present invention.

For diagnostic and therapeutic goals the described DNA segments may also be employed for expressing coded antigens, parts thereof or combinations thereof with alien antigens. Therein the DNA segments under aimed control of regulation sequences are introduced into pro- or eukaryotic target cells, tissues or multiple-cell organisms to stimulate these to produce the accordingly coded antigens, parts thereof or combinations thereof with alien antigens. Antigens can be detected via the reaction with anti-HIV-2 antibodies, more particularly from the sera of the respective patients. Antigens having longer open reading frames (>50 amino acids) lend themselves as well those which are subject to splicing processes on the RNA level and are only thus composed to form the longer open reading frames.

According to the invention further claimed are polypeptides originating from the cloned virus variant according to the invention to detect such antigens in the material under investigation which contain similar antigen determinants and thereby do immunologically cross-react. This is particularly suitable for the diagnosis of AIDS and pre-AIDS of virus carriers or asymptomatic virus carriers or virus products, respectively, which are derived from blood. Also the serological detection of the antibodies directed against these antigenic polypeptides as expression products of the virus claimed according to the invention becomes possible by employing conventional systems such as ELISA. The immunogenic polypeptides may be used as protective polypeptides as vaccines to cause protection against AIDS infections.

The polypeptides according to the invention are understood to include fragments which are intentionally obtained by means of gene-technological methods, starting from longer open reading frames as well as those obtained by proteolytic enzymes in the production bacterial strains or *in vitro* by the use of proteases.

The virus isolates according to the invention and the products derived therefrom may be combined with other isolates of the partial population HIV-2 in test systems, that is with those which are as far remote as possible in the described population level such as for example, the isolate HIV-2 ROD (Guyader, M. et al., 1987). Thereby it becomes possible sensitively to detect also populations of remote relationship in one test.

The virus variant according to the invention is highly different from the spectrum of the HIV-1 variants and have a closer molecular relationship to the HIV-2 virus described by Guyader, although they are distinguished therefrom to a significant extent (Figure 1). Also the biological properties are clearly distinguished from the described HIV-2 isolate. Thus, the variant according to the invention, for the effective *in vitro* replication, prefers cells which are derived from myeloidic lines. On the contrary, the virus poorly reproduces itself on lymphocytic lines.

A sample of the virus claimed according to the invention has been deposited in the form of its isolate at the European Collection of Animal Cell Cultures under the designation HIV D205 (V 87122304) according to the Budapest Treaty.

Figure 1 shows the restriction maps of the virus isolate according to the invention in comparison to known HIV sequences.

Figure 2 shows the partial nucleotide sequences of HIV-D205 (corresponding to clone HIV-2 A7.1 of Figure 2).

Figure 3 shows the sequence homology of HIV-2 D205,7 compared to the HIV/SIV group (gene level; nt/aa).

Figure 4 shows a nucleotide sequence comparison of HIV-2 D205 with HIV and SIV strains (in % homology).

Experimental results and characteristics of HIV-D205 are described in Kühnel, H. et al. (1989) Proc. Natl. Acad. Sci. 86, 4, 2383-2387.

The sequence of HIV-D205 shows a lot of so-called "open reading frames". Most of these reading frames can be related to *in vivo* expressed proteins/antigens by comparison of homologies to previously described HIV-viruses, by comparison of Western blots performed with HIV-D205 antigens derived from infected HUT78 or J937 cells and by probing with sera from the corresponding patients and reference sera.

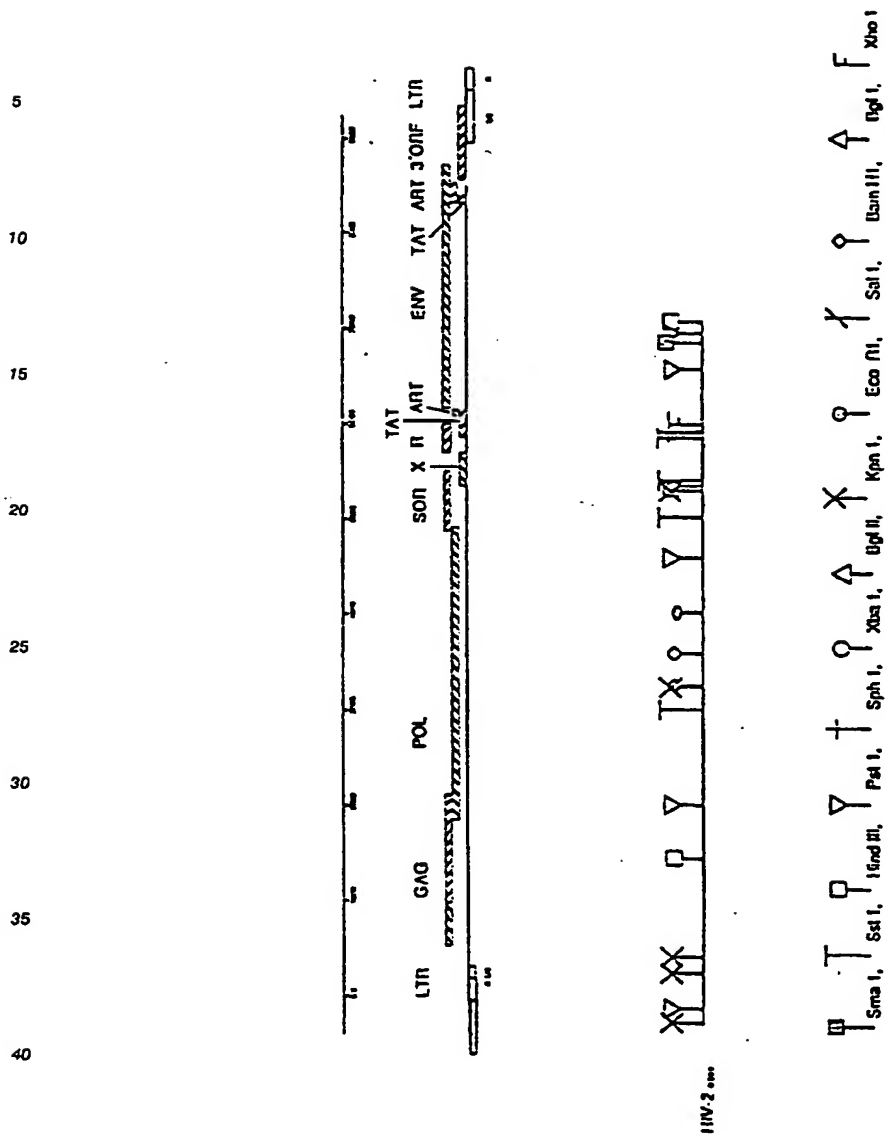
Other open reading frames are not identified on the level of their expressed antigens defined by function or antibody staining on Western Blot. However, they can be expressed under some circumstances *in vivo*. Other reading frames, even short ones, can be expressed as well in a way difficult to predict solely on the basis of nucleic acid sequencing data because of splicing processes.

Antigenic determinants on expressed proteins as they are important for the biological function, for target antigens in diagnostics or for immunization are spread all over the expressed linear protein sequence. Parts of these sequences can have more general anticenic properties than others as can be shown by peptide screening/ mapping for antigenic sites. These sites can be expressed as single epitopes or as continuous polypeptide or in a version of in vitro or synthetically spliced antigens. Antigenicity of the expressed products can be demonstrated by antigen fixation and blotting in the Western Blot assay. Constructions for antigen expression in E. coli can be done by using conventional techniques using synthetic genes, restriction fragments from cloned viral genome segments, trimming products thereof by using exonuclease or DNase I or by using sequence specific synthetic primers defining the desired 5' and 3' end of the fragment to be expressed together with appropriate restriction sites. These restriction sites can easily be used for ligation into a panel of expression vectors of different organisms like those derived from PLc24 (Remault et al. 1981 Gene 15, 81-83) with multicloning sites (pEX).

The expressed antigens were shown to specifically react with patients' sera. The p27(24) from gag of HIV-D205 react very sensitively with both typical HIV-1 sera and typical HIV-2 sera (see Kühnel et al).

Claims

1. A virus isolate HIV D205 (ECACC V 87122304).
2. DNA of the proviral partial sequences according to the following restriction endonuclease section-site characteristics, within the scope of the possible and conventional variation of errors, formed in establishing restriction maps.



3. cDNA and -fragments of the virus isolates according to claim 1.
4. Viral RNA and its fragments from virus isolates according to claim 1.
5. Recombinant DNA containing DNA pieces, starting from the virus isolates according to claim 1.
6. DNA or RNA of the virus isolates according to any one of the claims 1 to 4, wherein the DNA or RNA is present as hybide with complementary labelled DNA or RNA strands.
7. DNA according to any one of the claims 1 to 5, characterized in that it is complementary to viral DNA or parts thereof.

8. Nucleic acid strands in a modified or unmodified form which under stringent conditions hybridize with nucleic acids according to claims 2 to 7, and more specifically those nucleic acids which correspond to the highly variable regions of the HIV genom, more particularly in the range of the region coding the envelope protein.
- 5 9. Expression products of the virus isolates according to claim 1.
10. Expression products according to claim 1, characterized in that the proteins, peptides or fragments have been coded within the meaning of an open reading frame on the DNA according to claim 2.
- 10 11. A process for the in vitro detection of antibodies against expression products of the viruses according to claim 1, characterized in that the expression products or parts thereof of the viruses are detected by means of immunological methods.
- 15 12. The process according to claim 11, characterized in that the expression products are proteins, peptides or parts thereof which have been coded within the meaning of an open reading frame on the DNA according to claim 2 and are prepared by synthetic or biosynthetic processes.
- 20 13. The process according to claims 11 or 12, characterized in that previously a definite amount or a combination of expression products or parts thereof are fixed on microtiter plates, whereupon subsequently biological samples, diluted or undiluted, are contacted with the coated microtiter plates and after incubation and sequential washing steps can be identified by means of a detecting reagent or of labelled anti-HIV antibodies.
- 25 14. The process according to any one of claims 11 to 13, characterized in that filter strips and plastic strips or rods are used instead of microtiter plates, wherein the expression products of the viruses have been fixed at respective specific positions by isolated application of the different antigens.
- 30 15. The process according to claim 14, characterized in that the expression products or parts thereof are separated by gel electrophoresis and then transferred by blotting whereupon incubation with anti-HIV antibodies and the detection thereof are effected.
- 35 16. The process according to any one of claims 11 to 15, characterized in that the detection is effected on solid phase carriers to which the antigen determinants have been bonded. the solid phase carrier consisting of particles.
- 40 17. The process according to any one of claims 11 to 16, characterized in that the expression products are virus antigens derived from in vitro-infected cells, said anti-genes being contacted with biological test materials as antigens bonded to fixed cells, and that the subsequent antibody bonding can be determined with immunological detection reagents by means of an apparatus, for example with a cytofluorimeter, or visually.
- 45 18. The process according to any one of claims 11 to 17, characterized in that the antigens are determined by competitive ELISA.
- 50 19. A process for detecting HIV-related nucleic acids (DNA and RNA) in biological samples, cells and in isolated form by using the nucleic acids according to claims 2 to 7.
- 55 20. The process according to any one of claims 11 to 19, characterized in that the expression products are supplemented by materials which are related to other HIV variants, which, however, are distinguished in their biological properties from the materials of the isolates according to claim 1.
21. Immunogenic composition, containing expression products such as antigens, coded by the viruses of the virus isolates according to claim 1.
22. The immunogenic composition according to claim 21, characterized in that one antigen constitutes part of the total membrane antigen or is the total membrane antigen or a derivative thereof or a mixture of parts of the membrane antigens.

23. Antibodies, and more specifically monoclonal antibodies, against expression products of the virus isolates according to claim 1.

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24. Cells which have been transformed with nucleic acids according to any one of claims 2 to 7.

25. Cells which have been infected with virus isolates according to claim 1.

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Fig. 1

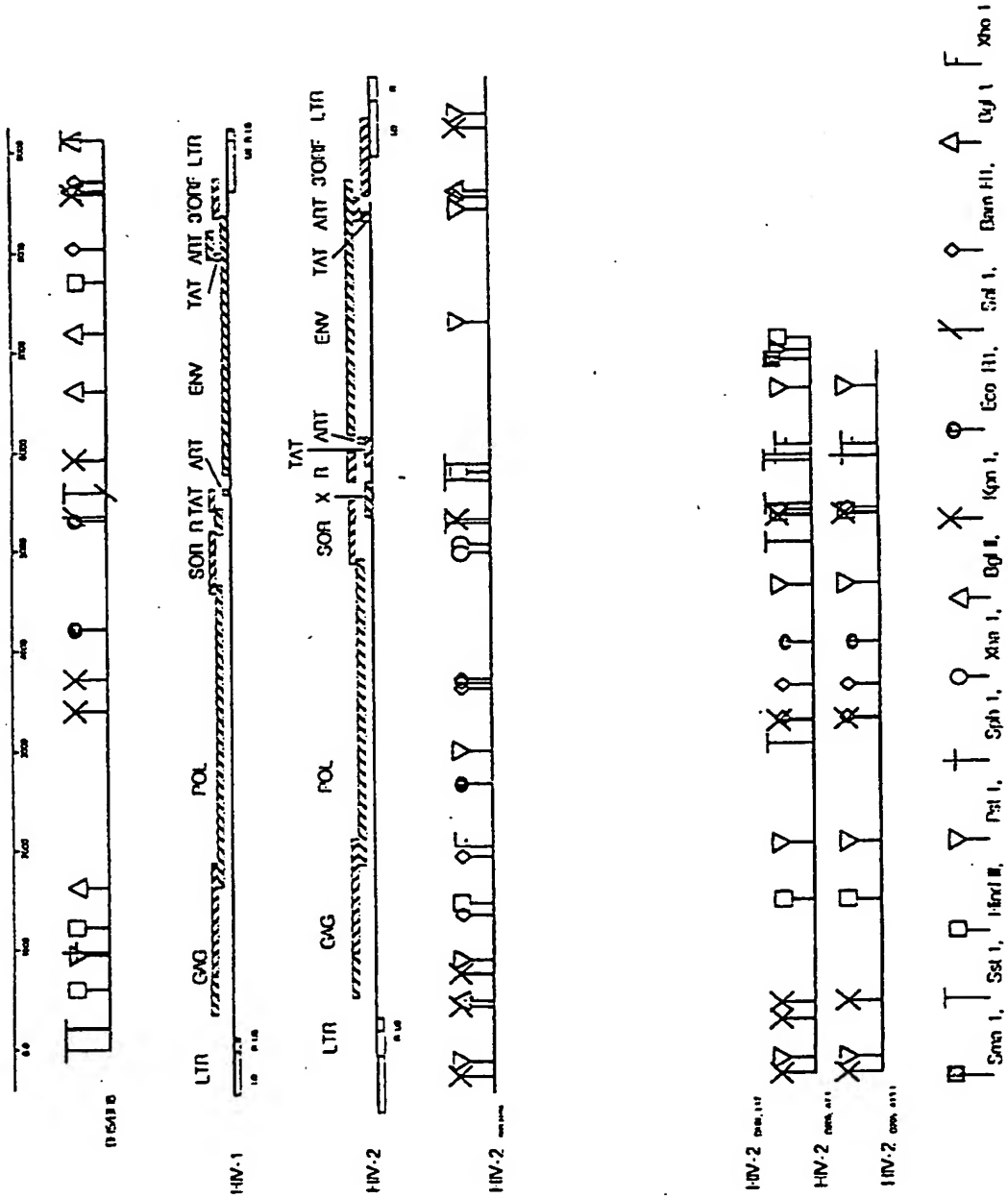


Fig. 2

Partial nucleotide sequences of HIV-D205
(corresponding to clone HIV-2 A7.1 of Fig. 2);

HIV-D205; corresponding to pcs. 8942-9255 in HIV-2 ROD; homology 71.6 %

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      10      20      30      40      50      60
TGG AAGGGAT GTATTATAGT GAGAGAAGAC ACAGAATATT AGACACATAT TTTGAGAATG

      70      80      90     100     110     120
AAGAAGGCAT TGTGTCTGGC TGGCAAAACT ATACTCATGG GCCAGGGATA AGGCATCCCA

     130     140     150     160     170     180
AATACTTTGG TTGGCTGTGG AAGCTGGTAC CAGTAGAGGT GCCAGCAGCG ACCCGAGAGG

     190     200     210     220     230     240
AGGAGGAAAC CCATTGCCTA ATGCACCCGG CACAGATCTC CTCATGGGAT GACATCCATG

     250     260     270     280     290     300
GGGAGACTCT TATCTGGCAG TTTGATTCCC TCCTGGCATA TGATTATGTG GCTTTCAATA

     310
GGTTTCCAGA AGAGTTT

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HIV-D205, corresponding to position 718-2510 in HIV-2 ROD; homology 78.6 %

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      10      20      30      40      50      60
AAAAAATTCT TAAAGTCTTA GCTCCATTAG TACCAACAGG GTCAGAAAT TTA AAAAGCC

      70      80      90     100     110     120
TTTTTAATAT CGTCTGCGTC ATTTTGTGCC TGCACGCAGA AGAGAAAGTG AAAGATACAG

     130     140     150     160     170     180
AGGAAGCAAA AAAGATAGCA CAGAGACATC TAGCGGCGGA CACAGAAAAA ATGCCAGCTA

     190     200     210     220     230     240
CAAATAAACC AACAGCACCA CCTAGCGGCG GAAATTATCC AGTGCAGCAA CTGGCTGGCA

     250     260     270     280     290     300
ACTACGTCCA CCTGCCGCTA AGCCCCCGAA CCTTAAATGC TTGGGTAAAG TTAGTAGAAG

     310     320     330     340     350     360
AAAAGAAGTT CGGGGCAGAA GTAGTACCAG GATTTCAGGC ACTATCAGAA GGATGCACCC

     370     380     390     400     410     420
CTTATGATAT AAATCAGATG CTAAATTGTG TAGGAGAACA TCAGGCAGCC ATGCAAATTA

     430     440     450     460     470     480
TTAGAGAAAT AATCAATGAG GAAGCAGCAG ACTGGGACCA GCAACACCCG TCACCAGGCC

```

Fig. 2

490	500	510	520	530	540
CAATGCCGGC	AGGACAACCTT	AGGGACCCAA	GAGGGTCAGA	TATAGCAGGA	ACCACCAGCA
550	560	570	580	590	600
CAGTAGAGGA	ACAGATACAG	TGGATGTACA	GGGCCCCAAA	TCCTGTCCCA	GTGGGAAACA
610	620	630	640	650	660
TTTATAGAAG	ATGGATTCAA	TTAGGATTGC	AGAAATGTGT	CCGAATGTAC	AATCCTACCA
670	680	690	700	710	720
ACATATTAGA	CATAAAGCAG	GGACCAAAGG	AGCCCTTCCA	AAGCTATGTA	GATAGATTCT
730	740	750	760	770	780
ACAAAAGCTT	ACGGGCAGAA	CAAACAGACC	CAGCAGTGAA	AAATTGGATG	ACACAAACAC
790	800	810	820	830	840
TGCTGATTCA	GAATGCTAAC	CCAGATTGCA	AGTTAGTGCT	TAAGGGCTTG	GGATGAATC
850	860	870	880	890	900
CCACCTTAGA	GGAAATGCTA	ACGGCCTGCC	AAGGGATAGG	AGGCCCAAGG	CAGAAGGCCA
910	920	930	940	950	960
GGCTAATGGC	CGAAGCCTTA	AAAGAGGCCC	TACACCTGC	ACCCATACCG	TTTGCTGCCG
970	980	990	1000	1010	1020
TTCAACAAAA	AGCAGGGAAG	AGAGGGACAG	TGACATGCTG	GAAGTGTGGC	AAACAGGGAC
1030	1040	1050	1060	1070	1080
ACACAGCCAG	GCAATGCAGG	GGCCCTAGAA	GACAGGGATG	CTGGAAATGT	GGAAAAACAG
1090	1100	1110	1120	1130	1140
GACACATCAT	GTCAAAATGC	CCAGAAAGAC	AGGCGGGTTT	TTTAGGGTTA	GGACCCTGGG
1150	1160	1170	1180	1190	1200
GAAAGAAGCC	TCGCAACTTC	CCCATGACCC	AAGTGCCTCA	GGGAGTGACA	CCATCTGCAC
1210	1220	1230	1240	1250	1260
CCCCGATGAA	CCCAGCAGAG	GGCATGACAC	CTCGGGGGGC	GACACCATCT	GCGCCCCCTG
1270	1280	1290	1300	1310	1320
CAGATCCAGC	AGTGGAGATG	CTGAAAAGTT	ACATGCAGAT	GGGGAGACAA	CAGAGAGAGA
1330	1340	1350	1360	1370	1380
GCCGAGAGAG	ACCCTACAAG	GAGGTGACAG	AGGATTGCT	GCACCTCAAT	TCTCTCTTG
1390	1400	1410	1420	1430	1440
GAGAAGACCA	GTAGTCAAAG	CATGTATCGA	GGGTCACTCA	GTAGAAGTAT	TACTAGACAC
1450	1460	1470	1480	1490	1500
AGGAGTTGAC	GACTCAATAG	TAGCAGGGAT	AGAATTAGGT	AGCAATTACA	CCCCAAAAT
1510	1520	1530	1540	1550	1560
AGTAGGAGGG	ATAGGAGGGT	TCATAAATAC	CAAGAATAC	AAAGATGTAG	AAATAGAAAT
1570	1580	1590	1600	1610	1620
AGTGGGAAAA	AGAGTAAGGG	CAACTATAAT	GACAGGAGAT	ACCCCAATAA	ACATTTTGG

Fig. 2

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      1630      1640      1650      1660      1670      1680
CAGAAATATT TTAATACCT TGGGCATGAC TTAAATTTC CCAGTGGCAA AGGTAGAACC

      1690      1700      1710      1720      1730      1740
AGTAAAAGTT GAGTTAAAC CTGGAAGA TGGGCCAAG ATCAGACAAT GGCCTCTATC

      1750      1760      1770      1780      1790
CAGGGAAAAG ATACTAGCCC TCAAAGAAAT CTGTGAAAAA ATGGAAAAGG

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HIV-D205, corresponding to position 2877-7293 in HIV-2ROD; homology 75.1 %.

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      10      20      30      40      50      60
AGGTATTAGA TCCTTTTGA AAGGCCAACA GCGATGTCAT TATAATTGAG TACATGGATG

      70      80      90      100     110     120
ACATCCTTAT AGCAAGTGAC AGAAGTGATC TGGAGCACGA CAGGGTAGTG TCCCAACTAA

      130     140     150     160     170     180
AAGAGTTATT AAATGACATG GGATTCTCTA CCCCAGAAGA AAAGTTCCAA AAAGACCCTC

      190     200     210     220     230     240
CGTTCAAATG GATGGGTTAT GAGCTCTGGC CAAAAAGTG GAAACTGCAA AAATACAAC

      250     260     270     280     290     300
TGCCAGAAA AGAAGTTTGG ACAGTGAATG CAATTCAAAA ACTGGTAGGA GTATTAACT

      310     320     330     340     350     360
GGGCAGCTCA ACTCTTTCCT GGAATTAAGA CAAGGCACAT ATGCAAACTA ATTAGGGGAA

      370     380     390     400     410     420
AGATGACCCT AACAGAAGAA GTACAGTGGA CAGAACTAGC AGAAGCAGAG CTACAGGAGA

      430     440     450     460     470     480
ATAAAATCAT CTTAGAACAG GAACAAGAAG GATCCTACTA CAAGGAAAGG GTACCGCTAG

      490     500     510     520     530     540
AAGCAACAGT ACAGAAAAAC CTAGCAAATC AGTGGACATA CAAATTCAT CAGGGAAATA

      550     560     570     580     590     600
AAGTCCTAAA AGTAGGAAAA TATGCAAAGG TTAAAAACAC GCACACCAAC GGGGTAAGAC

      610     620     630     640     650     660
TACTGGCACA TGTAGTTCAG AAAATAGGCA AAGAAGCCCT AGTCATCTGG GGAGAGATAC

      670     680     690     700     710     720
CAGTGTTCCA TCTGCCAGTA GAAAGAGAGA CATGGGACCA GTGGTGGACA GATTACTGGC

      730     740     750     760     770     780
AAGTAACCTG GATCCCAGAG TGGGACTTTG TCTCGACCCC ACCATTAATA AGACTAGCCT

      790     800     810     820     830     840
ACAACCTAGT CAAAGACCCC CTAGAAGGGA GAGAAACCTA CTACACAGAT GGGTCCTGCA

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Fig. 2

850	860	870	880	890	900
ATAGAACCTC	AAAGGAAGGA	AAAGCAGGAT	ATGTCACTGA	CAGGGGAAAA	GATAAGGTTA
910	920	930	940	950	960
AAGTGTTAGA	ACAGACAACA	AACCAACAAG	CAGAACTTGA	AGCATTGCA	TTAGCATTAA
970	980	990	1000	1010	1020
CAGACTCAGA	ACCACAAGTT	AACATCATAG	TAGATTCACA	ATATGTCATG	GGAATAATAG
1030	1040	1050	1060	1070	1080
CTGCACAGCC	AACAGAAACA	GAATCACCAA	TAGTAGCAAA	AATAATTGAA	GAAATGATCA
1090	1100	1110	1120	1130	1140
AAAAAGAGGC	AGTATATGTA	GGATGGGTAC	CAGCTCACAA	GGGACTGGGT	GGTAATCAGG
1150	1160	1170	1180	1190	1200
AAGTAGACCA	CCTAGTAAGT	CAAGGAATCA	GACAGGTCTT	GTTCTAGAA	AAAATAGAAC
1210	1220	1230	1240	1250	1260
CAGCCCAGGA	AGAGCATGAA	AAATATCATG	GCAATGTAAA	AGAACTGGTC	CATAAATTCG
1270	1280	1290	1300	1310	1320
GAATTCCACA	ATTAGTGGCA	AAACAGATAG	TAAATTCCTG	TGATAAATGC	CAACAAAAAG
1330	1340	1350	1360	1370	1380
GGGAAGCTAT	TCATGGACAG	GTAAATGCAG	ACCTAGGGAC	ATGGCAGATG	GACTGTACAC
1390	1400	1410	1420	1430	1440
ATTTAGAAGG	AAAAATTATA	ATAGTGGCAG	TCCATGTAGC	CAGTGGGTTT	ATAGAAGCAG
1450	1460	1470	1480	1490	1500
AGGTAATACC	CCAAGAGACA	GGAAGACAGA	CAGCTCTCTT	CCTACTAAAG	TTGGCCAGCA
1510	1520	1530	1540	1550	1560
GATGGCCTAT	CACACACCTA	CACACAGACA	ACGGTGCCAA	CTTCACCTCA	CCAAGTGTA
1570	1580	1590	1600	1610	1620
AGATGGTAGC	CTGGTGGGTA	GGAATAGAAC	AAACTTTTGG	AGTACCCTAT	AACCCACAAA
1630	1640	1650	1660	1670	1680
GTCAAGGAGT	AGTGGAAGCA	ATGAACCATC	ACCTGAAAAA	TCAAATAGAC	AGACTCAGAG
1690	1700	1710	1720	1730	1740
ACCAAGCAGT	ATCAATAGAG	ACAGTTGTAC	TAAATGGCAAC	TCACTGCATG	AATTTTAAAA
1750	1760	1770	1780	1790	1800
GAAGGGGAGG	AATAGGGGAT	ATGACCCCTG	CAGAAAAGACT	AGTTAACATG	ATAACCACAG
1810	1820	1830	1840	1850	1860
AGCAAGAAAT	ACAGTTCTTC	CAAGCAAAAA	ATTTAAAAAT	TCAAAATTTT	CAGGTCTATT
1870	1880	1890	1900	1910	1920
ACAGAGAAGG	CAGAGATCAA	CTCTGGGAAG	GACCTGGTGA	ACTATTCTGC	AAAGGGGAAG
1930	1940	1950	1960	1970	1980
GAGCAGTCAT	CATAAAGGTA	GGGACAGAAA	TCAAAGTAGT	ACCCAGGAGA	AAAGCAAAAA

Fig. 2

1990	2000	2010	2020	2030	2040
TTATAAGGCA	CTATGGAGGA	GGAAAAGGAT	TGGATTGTAG	TGCCGACATG	GAGGATACCA
2050	2060	2070	2080	2090	2100
GGCAGGCTAG	AGAGATGGCA	CAGTCTGATT	AAGTATCTTA	AGTATAGAAC	AGGAGAGTTG
2110	2120	2130	2140	2150	2160
CAACAGGTCT	CTTATGTCCC	TCACCACAAG	GTAGGATGGG	CTTGGTGGAC	TTGCAGTAGA
2170	2180	2190	2200	2210	2220
ATAATATTC	CCCTAAACAA	AGGAGCATGG	CTAGAAGTCC	AAGGATATTG	GAACCTAAC
2230	2240	2250	2260	2270	2280
CCAGAAAGGG	GATTCTTGAG	CTCCTATGCT	GTAAAGACTAA	CATGGTATGA	GAGGAACCTT
2290	2300	2310	2320	2330	2340
TATACAGATG	TAACACCTGA	TGTGGCAGAC	CAGCTACTGC	ATGGGTCTTA	TTTCTCTTGC
2350	2360	2370	2380	2390	2400
TTTTTCAGCCA	ATGAAGTAAG	GAGAGCCATC	AGGGGAGAAA	AGATATTGTC	CTACTGCAAC
2410	2420	2430	2440	2450	2460
TATCCATCAG	CTCAGGAAGG	GCAGGTACCA	AGCTTACAGT	TTCTAGCCCT	AAGGGTCGTA
2470	2480	2490	2500	2510	2520
CAGGAAGGAA	AAAATGGATC	CCAGGGAGAG	AGTGCCACCA	GGAAACAGCG	ACGAAGAAAC
2530	2540	2550	2560	2570	2580
AGTAGGAGAA	GCATTGCTT	GGCTAGAAAG	AACAATAACA	GAGCTCAACA	GGGTAGCGGT
2590	2600	2610	2620	2630	2640
CAACCATTTG	CCCCGAGAAC	TTATTTTCCA	GGTCTGGCAG	AGGTCTTGGG	CATACTGGCG
2650	2660	2670	2680	2690	2700
TGAGGAACAG	GGCATGTCAA	TTAGCTATAC	CAAAATATAGA	TACTTGTTGC	TAATGCAGAA
2710	2720	2730	2740	2750	2760
AGCAATGTTT	GTGCACTATA	CAAAGGGCTG	TAGGTGCCTG	CAGGAGGGCC	ATGGGCCAGG
2770	2780	2790	2800	2810	2820
GGGATNGAGA	TCAGGACCTC	CTCCTCCTCC	TCCCCCAGGC	CTGGCCTAAT	GGCAGAAGCA
2830	2840	2850	2860	2870	2880
GGCCCAAGAGA	TCCCTCCAGA	GAACGAGAAC	CCACAAAGAG	AACCGTGGGA	AGAGTGGATA
2890	2900	2910	2920	2930	2940
GGGGAGATCC	TGGAGGAAAT	AAAGCAAGAA	GCCTTAAAGC	ATTTTGATCC	TGGCTTGCTA
2950	2960	2970	2980	2990	3000
ACTGCGCTTG	GTAACTTTAT	CTACAGTAGG	CATGGAGATA	CCCTTGACAGG	AGCAGGAGAG
3010	3020	3030	3040	3050	3060
CTCATTAATA	TCCTCCAACG	AGCNCTCTTC	CTCCACTTCA	GAGCCGGTTG	TCAACACTCA
3070	3080	3090	3100	3110	3120
AGGATTGGAC	AATCAGGGGG	AGGAAATCCT	CTCTCAACTA	TACCGCCCCC	TTAAGGCATG

Fig. 2

3130	3140	3150	3160	3170	3180
CGATAATACA	TGCTACTGTA	AGAAATGCTG	CTACCATTGC	CAGCTTTGTT	TTCTTAAAAA
3190	3200	3210	3220	3230	3240
GGGTCTTGGG	ATATGTTATG	ACCGCTCGAG	AAGGAGATCT	GCAAAAGAG	CTAAGACTAC
3250	3260	3270	3280	3290	3300
TGCACCTTCT	GCACCAGACA	AGTGAGTATG	GCATATTTTA	GCAGCCGCCT	GCCTATTGCG
3310	3320	3330	3340	3350	3360
CTCCTGCTTA	TAGGTATCAG	TGGGTTTGTA	TGTAAACAAT	ATGTTACTGT	CTTCTATGGC
3370	3380	3390	3400	3410	3420
ATACCCGCAT	GGAGGAACGC	AACAGTCCCC	CTCATTGTGT	CAACCACAA	CAGAGACACC
3430	3440	3450	3460	3470	3480
TGGGGAACTG	TACAGTGTCT	CCCAGACAAT	GGTGACTACA	CTGAGATCAG	GCTAAACATA
3490	3500	3510	3520	3530	3540
ACAGAGGCTT	TTGATGCATG	GGATAATACA	GTGACACAAC	AGGCAGTAGA	TGATGTGTGG
3550	3560	3570	3580	3590	3600
AGACTCTTTG	AAACCTCCAT	AAAACCATGT	GTCAAATAA	CCCCACTGTG	TGTGGCAATG
3610	3620	3630	3640	3650	3660
AACTGTAGTA	AAACCGAAAC	AAACCCAGGG	AATGCCAGTA	GTACTACCAC	CACTAGCCT
3670	3680	3690	3700	3710	3720
ACTACCACCT	CTCGTGGGCT	GAAAACGATT	AACGAAACAG	ACCCATGCAT	AAAAATGAC
3730	3740	3750	3760	3770	3780
AGCTGCACAG	GA CTAGGAGA	AGAGGAAATA	ATGCAATGTA	ATTTTAGTAT	GACGGGACTA
3790	3800	3810	3820	3830	3840
AGAAGAGATG	AGCTAAACA	ATATAAGAC	ACCTGGTACT	CAGAAGATTT	AGAGTGTAAT
3850	3860	3870	3880	3890	3900
AATACCAGGA	AGTAATACCA	GCAGTGCTAT	ATAAGAACCT	GCAACACAAC	AATTATCCAA
3910	3920	3930	3940	3950	3960
GAGTCATGTG	ACAAACATTA	TTGGGACAGC	TTAAGGTTTA	GGTATTGTGC	TCCCCCGGGG
3970	3980	3990	4000	4010	4020
TTTTTTCTAC	TAAGATGTAA	TGATACCAAC	TATTCAGGCT	TCATGCCCAA	CTGCAGTAAG
4030	4040	4050	4060	4070	4080
GTAGTAGCGT	CCTCCTGCAC	AAGAATGATG	GAAACACAGT	CCTCTACATG	GTTTGGCTTC
4090	4100	4110	4120	4130	4140
AATGGTACAA	GGGCAGAGAA	CAGGACATAT	ATATATTGGC	ATGAAAAAGA	CAATAGGACC
4150	4160	4170	4180	4190	4200
ATCATAAGCT	TAAATACATA	CTATAATTTG	TCAATACACT	GTAAGAGGCC	AGGAAAACAG
4210	4220	4230	4240	4250	4260
ACGGTTGTAC	CAATAAGAAC	CGTGTCAGGA	CTACTTTTCC	ATTCACAGCC	TATCAATAAG

Fig. 2

4270	4280	4290	4300	4310	4320
AGACCCAGAC	AAGCTTGGTG	CTGGTTTAAG	GGAAACTGGA	CAGAAGCCAT	AAAGGAGGTG
4330	4340	4350	4360	4370	4380
AAAAGGACCA	TCATAAAACA	TCCCAGGTAT	AAAGGAGGTG	CAAAAAATAT	CACAAGCGTA
4390	4400	4410			
AAGTTAGTAT	CAGAACATGG	AAAAGGTTCA	GATC		

Fig. 3

Sequence homology of HIV-2_{D205,7} compared to the HIV/SIV group (gene level; nt / aa)

HIV-2 _{D205,7}									
gene	position	HIV-2 _{ROD}	HIV-2 _{NIHZ}	HIV-2 _{D194}	SIV _{MAC}	SIV _{AGM}	HIV-1 _{BRU}		
gag	720-1026	80.5 / 85.6							
gag	1860-2114	83.1 / 77.6							
pol	1859-2510	80.2 / 72.5							
pol	2077-4948	78.3 / 83.5							
protease	2084-2301	84.0 / 81.0	83.0 / 84.8	84.8 / 86.8	76.3 / 83.8	57.8 / 47.1	60.4 / 48.5		
vif	4869-5516	72.0 / 68.5	70.9 / 67.9	72.4 / 66.5	71.8 / 60.6	53.8 / 34.7	47.9 / 33.0		
vpx	5344-5682	76.1 / 74.1	73.5 / 68.1	74.6 / 77.9	75.2 / 77.0	50.8 / 34.7			
vpr	5682-5999	70.8 / 69.8	77.7 / 69.8	74.2 / 59.4	78.3 / 76.4		51.9 / 47.3		
latex1	5845-6140	78.4 / 66.3	79.1 / 68.4	74.7 / 63.3	81.1 / 66.3	33.1 / 38.1	33.6 / 34.0		
revex1	6071-6140	67.1 / 61.9	68.6 / 60.9	67.1 / 52.2	70.3 / 60.9	45.5 / 28.6	38.2 / 40.4		
nei	8557-9255	72.1 / 69.5							
env	6147-7293	70.0 / 67.0							

Fig. 4

Nucleotide sequence comparison of HIV-2_{D205} with HIV and SIV strains (in % homology)

HIV-2 _{D205}						
position	HIV-2 _{ROD}	HIV-2 _{NIH}	HIV-2 _{D194}	SIV _{MAC}	SIV _{AGM}	HIV-1 _{BRU}
8942-9255	71.6	77.0	68.8	66.4	56.3	54.7
718-1825	80.5	80.8	80.3	79.1	65.1	63.8
1059-2510	80.2	74.6	75.0	70.8	55.0	56.9
2877-7293	75.1	74.8	75.4	74.0	50.0	54.6
Total	75.9	75.9	75.9	75.0	50.9	56.4



European Patent
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EUROPEAN SEARCH REPORT

Application Number
EP 95 10 0149

DOCUMENTS CONSIDERED TO BE RELEVANT			
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int.Cl.4)
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The present search report has been drawn up for all claims			
Place of search THE HAGUE		Date of completion of the search 17 February 1995	Examiner Cupido, M
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X : particularly relevant if taken alone Y : particularly relevant if combined with another document of the same category A : technological background O : non-written disclosure F : intermediate document		T : theory or principle underlying the invention E : earlier patent document, not published on, or after the filing date D : document cited in the application L : document cited for other reasons @ : member of the same patent family, corresponding document	